ANCHOR DIAGNOSTICS

SpeedPURE® Go Kit (48)
SpeedPURE® Multi Kit (96)
SpeedPURE® Flex Kit (384)

Instructions for Use

SpeedPURE® Kits

RUO

Nucleic acid extraction kits

For Research Use Only

RUO For Research Use Only

REF AP03400S: SpeedPURE® Go Kit (48)
AP03400M: SpeedPURE® Multi Kit (96)
AP03400L: SpeedPURE® Flex Kit (384)

SpeedPURE® Go Kit: 48
SpeedPURE® Multi Kit: 96
SpeedPURE® Flex Kit: 384

AP03490-EN, 2026-07-28

QG AP03491S-EN, 2026-07-24
AP03491M-EN, 2026-07-24
AP03491L-EN, 2026-07-24

+15°C to +25°C

ANCHOR Diagnostics GmbH
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Instructions for Use – SpeedPURE® Kits

compatible with

ANCHOR BEAD 32 (Mat. No. AP03432)

Instructions for Use – SpeedPURE® Kits

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2 ▸ Application

The SpeedPURE® Kits provide all reagents required for the automated extraction of nucleic acids from various biological sample materials in combination with the ANCHOR BEAD 32 device.

3 ▸ Test Principle

The SpeedPURE® Kits contain four different buffers, an enzyme (Proteinase K) and magnetic beads. For the SpeedPURE® Go Kit (48) and SpeedPURE® Multi Kit (96) configuration, the buffers and magnetic beads are provided in prefilled ready-to-use reagent vessels.
The SpeedPURE® Flex Kit (384) configuration provides maximum flexibility, enabling users to tailor reagent usage to their needs and optimize workflows for different sample throughputs.
All kits are designed to be used in combination with the ANCHOR BEAD 32 device utilizing the pre-installed programs. The optimized buffer composition ensures efficient nucleic acid purification. The Lysis Buffer releases nucleic acids, which then bind to the surface of magnetic beads. Subsequent washes with Wash Buffers I and II remove impurities, and the purified nucleic acids are finally eluted from the beads into the Elution Buffer. The resulting nucleic acids are suitable for downstream applications such as real-time PCR (e.g., using ANCHOR PCR Kits).
For the preparation of nucleic acid from stool and whole blood additional pre-treatment is recommended using the SpeedPURE® Dilution Buffer or the SpeedPURE® Pre-Lysis Buffer, respectively. For detailed instructions, please refer to sections 5.5 or 5.6, respectively.

4 ▸ Kit Components

4.1 ▸ SpeedPURE® Go Kit

Kit Component	Number / Volume
SpeedPURE® Go Cartridge	48
Proteinase K	1 x 1 mL
SpeedPURE® Magnetic Rod Covers*	6 x 2 covers
Quick Guide	1

*additional SpeedPURE® Magnetic Rod Covers can be purchased separately (50 pcs., Mat no. AP03462)

SpeedPURE® Go Cartridge Contents		
Well	**Buffer**	**Volume (µL)**
1	Lysis Buffer	600
2	Wash Buffer I	600
3	Wash Buffer II	600
4	Magnetic Beads	80
5	(empty)	0
6	Elution Buffer*	90

*contains sodium azide as a preservative component

4.2 ▸ SpeedPURE® Multi Kit

Kit Component	Number / Volume
SpeedPURE® Multi Plate	6
Proteinase K	2 x 1 mL
SpeedPURE® Magnetic Rod Covers*	6 x 2 covers
Quick Guide	1

*additional SpeedPURE® Magnetic Rod Covers can be purchased separately (50 pcs., Mat no. AP03462)

SpeedPURE® Multi Plate Contents		
Wells	**Buffer**	**Volume (µL)**
1 & 7	Lysis Buffer	600
2 & 8	Wash Buffer I	600
3 & 9	Wash Buffer II	600
4 & 10	Magnetic Beads	80
5 & 11	(empty)	0
6 & 12	Elution Buffer*	90

*contains sodium azide as a preservative component

4.3 ▸ SpeedPURE® Flex Kit

Kit Component	Number / Volume
Lysis Buffer	1 x 250 mL
Wash Buffer I	1 x 250 mL
Wash Buffer II	1 x 250 mL
Magnetic Beads	1 x 32 mL
Elution Buffer*	1 x 36 mL
Proteinase K	8 x 1 mL
Quick Guide	1

*contains sodium azide as a preservative component

When used in combination with ANCHOR BEAD 32, we recommend to use SpeedPURE® 96-well Deep-well plates (Mat. No. AP03461) and SpeedPURE® Magnetic Rod Covers (Mat No. AP03462).

5 ▸ Assay Procedure

Please refer to Quick Guide cards included in the kit boxes for detailed instructions (also see chapters 5.2, 5.3, and 5.4). The SpeedPURE® Kits can be used with various biological samples. It is recommended that nucleic acid extraction and subsequent analysis using the purified nucleic acids are properly controlled (e.g., with appropriate extraction and/or amplification controls).

5.1 ▸ Proteinase K + Internal Control Mix

The NA isolation using SpeedPURE® reagents requires 20 µL of Proteinase K for each sample.
If an ANCHOR PCR Kit is used as a downstream application, the Internal Control (IC) can be used as an extraction control. In this case, add 9 µL IC to the sample. If other downstream applications featuring an IC are used, please refer to the relevant IFU for more details.
For more than 8 samples, we recommend preparing a Proteinase K + IC Mix with 10% additional volume of each component. Add 29 µL of this Proteinase K + IC Mix to the sample/Lysis Buffer mix.

5.2 ▸ SpeedPURE® Go Kit

ANCHOR

SpeedPURE® Go Kit (48) (RUO)

1. Turn on the ANCHOR BEAD 32 and log in
2. Invert each Go Cartridge 3 times to fully resuspend the beads
3. Briefly flick the Go Cartridge downward by hand (3 times) to collect any liquid at the bottom of the wells
4. Insert one Go Cartridge per sample into the Go Cartridge Holder and remove the seal (Fig. 1)
5. Add 200 µL (or 400 µL, depending on protocol) of sample directly into the Lysis Buffer (Fig 2; first well, blue). **Do not mix!**
6. Addition of Proteinase K and Internal Control (IC)
   a. IC included as extraction control: add 20 µL of Proteinase K and 9 µL of IC to the sample/Lysis Buffer mix in the first well. **Do not mix!**
   b. IC **not** included as extraction control: add 20 µL of Proteinase K to the sample/Lysis Buffer mix in the first well. **Do not mix!**
7. Open the ANCHOR BEAD 32 door and securely place holder with Go Cartridges into the instrument with the cartridge labels facing the front
8. Insert the SpeedPURE® Magnetic Rod Covers – always use two rod covers per stand (two or four in total) to ensure proper protection of the magnetic rods (Fig. 3)

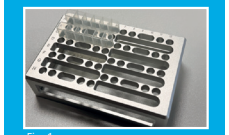
Fig. 1

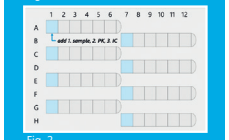

Fig. 2

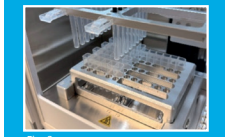
Fig. 3

ANCHOR

SpeedPURE® Go Kit (48) (RUO)

9. Ensure all magnetic rod covers click into place and are inserted firmly
10. On the display select Run (upper left corner)
11. Choose the desired protocol (Fig. 4)
    - SpeedPURE_200 for 200 µL sample input
    - SpeedPURE_400 for 400 µL sample input
12. Select Run (lower right corner) to start the sample preparation
13. After the run is complete:
    - Remove and discard magnetic rod covers
    - Remove Go Cartridge holder from instrument and collect eluate from well position 6 (Fig. 5)
    - Discard used Go Cartridge

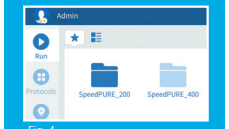

Fig. 4

Fig. 5

REF AP03400S QG AP03491S-EN, 2026-07-24

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5.3 ▸ SpeedPURE® Multi Kit

ANCHOR

SpeedPURE® Multi Kit (96)
(RUO)

1. Turn on the ANCHOR BEAD 32 and log in
2. Invert each Multi Plate 3 times to resuspend the beads
3. Briefly flick the filled plate downward by hand (3 times) to collect any liquid at the bottom of the wells
4. Remove the seal of each Multi Plate
5. Add 200 µL (or 400 µL, depending on protocol) of sample directly into the Lysis Buffer (Fig. 1; first well, blue). **Do not mix!**
6. Addition of Proteinase K and Internal Control (IC)
   a. IC included as extraction control: add 20 µL of Proteinase K and 9 µL of IC to the sample/Lysis Buffer mix in the first well. **Do not mix!**
   b. IC **not** included as extraction control: add 20 µL of Proteinase K to the sample/Lysis Buffer mix in the first well. **Do not mix!**
7. Open the ANCHOR BEAD 32 door and place Multi Plates into the instrument (Fig. 2) Ensure the Multi Plates are securely positioned with the plate label facing the front of the instrument
8. Insert the SpeedPURE® Magnetic Rod Covers – always use two rod covers per plate (two or four in total) to ensure proper protection of the magnetic rods (Fig. 3)

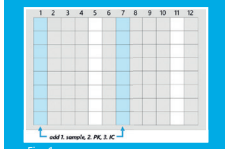

Fig. 1

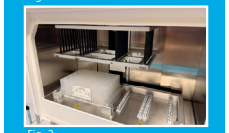

Fig. 2

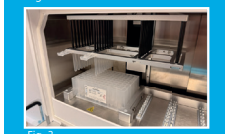

Fig. 3

ANCHOR

SpeedPURE® Multi Kit (96)
(RUO)

9. Ensure all magnetic rod covers click into place and are inserted firmly
10. On the display select Run (upper left corner)
11. Choose the desired protocol (Fig. 4)
    - SpeedPURE_200 for 200 µL sample input
    - SpeedPURE_400 for 400 µL sample input
12. Select Run (lower right corner) to start the sample preparation
13. After the run is complete:
    - Remove and discard magnetic rod covers
    - Remove Multi Plates from instrument and collect eluates from columns 6 and 12 (Fig. 5)
    - Discard used Multi Plates

Fig. 4

Fig. 5

REF AP03400M QG AP03491M-EN, 2026-07-24

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5.4 ▸ SpeedPURE® Flex Kit

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SpeedPURE® Flex Kit (384)
(RUO)

1. Turn on the ANCHOR BEAD 32 and log in
2. Prepare Deep Well Plate according to table A
3. **Important! Mix magnetic beads well before pipetting!**
4. Starting from the upper left, only prepare as many rows with reagents as needed for your samples (example shown for 10 samples, Fig. 1)
5. Add 200 µL (or 400 µL, depending on protocol) of sample directly into the Lysis Buffer (Fig. 1; first well, blue). **Do not mix!**
6. Addition of Proteinase K and Internal Control (IC):
   a. IC included as extraction control: add 20 µL of Proteinase K and 9 µL of IC to the sample/Lysis Buffer mix in the first well. **Do not mix!**
   b. IC **not** included as extraction control: add 20 µL of Proteinase K to the sample/Lysis Buffer mix in the first well. **Do not mix!**
7. Open the ANCHOR BEAD 32 door and place Deep Well Plates into the instrument (Fig. 2) Ensure the Deep Well Plates are securely positioned with the blue lysis buffer on the left
8. Insert the SpeedPURE® Magnetic Rod Covers – always use two rod covers per plate (two or four in total) to ensure proper protection of the magnetic rods (Fig. 3)

Columns	Reagent	Volume [µL]
1, 7	Lysis Buffer (L)	600
2, 8	Wash Buffer I (WI)	600
3, 9	Wash Buffer II (WII)	600
4, 10	Magnetic Beads (B)	80
5, 11	None (empty)	n/a
6, 12	Elution Buffer (E)	90

Table A

Fig. 1

Fig. 2

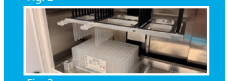

Fig. 3

ANCHOR

SpeedPURE® Flex Kit (384)
(RUO)

9. Ensure all magnetic rod covers click into place and are inserted firmly
10. On the display select Run (upper left corner)
11. Choose the desired protocol (Fig. 4)
    - SpeedPURE_200 for 200 µL sample input
    - SpeedPURE_400 for 400 µL sample input
12. Select Run (lower right corner) to start the sample preparation
13. After the run is complete:
    - Remove and discard magnetic rod covers
    - Remove Deep Well Plates from instrument and collect eluates from columns 6 and 12 (Fig. 5)
    - Discard used Deep Well Plates

Fig. 4

Fig. 5

REF AP03400L QG AP03491L-EN, 2026-07-24

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5.5 ▸ Handling of Stool Samples

For optimized performance, we recommend suspending stool samples in SpeedPURE® Dilution Buffer (Mat. No. AP03441):

- Prepare reaction tube with 1,000 µL of SpeedPURE® Dilution Buffer
- Transfer approx. 100 mg (pea-sized amount) or 100 µL of stool sample into reaction tube containing SpeedPURE® Dilution Buffer
- Pulse-vortex sample three times at maximum speed
- For best nucleic acid yields, we recommend different centrifugation protocols depending on the pathogen type.
  - Centrifuge samples for 5 min at 4,500 g for the isolation of viruses
  - Centrifuge samples for 1 min at 500 g for the isolation of bacteria/fungi
- Add 200 µL of the supernatant to the first well (lysis buffer, blue) of the SpeedPURE® kit according to chapters 5.2, 5.3, and 5.4

5.6 ▸ Handling of Whole Blood Samples

For optimized performance, we recommend pre-treating whole blood samples using the SpeedPURE® Pre-Lysis Buffer (Mat. No. AP03442):

- Invert sample in collection tube three times
- Prepare reaction tube with 220 µL of SpeedPURE® Pre-Lysis Buffer
- Add 220 µL of whole blood to the SpeedPURE® Pre-Lysis Buffer
- Pulse-vortex sample three times at maximum speed
- **Briefly** centrifuge mixture to prevent spillage
- Add 400 µL of this mixture to the lysis reaction of the SpeedPURE® kit according to chapters 5.2, 5.3, and 5.4.

6 ▸ Kit Storage

The kits are designed to be stored at room temperature (+15°C to +25°C). Please store all kit components inside the kit boxes protected from direct sunlight. Do not freeze any of the components. Always store boxes upright with the main label facing up.

7 ▸ Material Required but Not Provided

7.1 ▸ General Lab Equipment

- ANCHOR BEAD 32 nucleic acid extraction device (Mat. No. AP03432)
- Micropipettes (suitable volume range)
- Multi-channel pipettes for convenient filling of deep-well plates when using SpeedPURE® Flex Kits
- Single use trough for multi-channel pipettes
- Single-use pipette filter tips
- 1.5 mL or 2 mL reaction tubes (for Proteinase K + IC mix setup)
- Single-use gloves (powder-free)

7.2 ▸ SpeedPURE® Equipment and Consumabes

The following consumables are recommended for the use with the ANCHOR BEAD 32:

Mat. no.	Item	Pack Size
AP03431	SpeedPURE® Go Cartridge Holder	1
AP03462	SpeedPURE® Magnetic Rod Covers	50
AP03461	SpeedPURE® Deep-Well Plates (for use with SpeedPURE® Flex Kit)	10
AP03441	SpeedPURE® Dilution Buffer	100 extractions
AP03442	SpeedPURE® Pre-Lysis Buffer	500 extractions

8 ▸ Limitations

- Strict compliance with the instructions for use is required for optimal results.
- Following good laboratory practices is crucial for successful use of the product.
- Appropriate handling of the reagents is essential to avoid contamination or impurities.

9 ▸ Warnings and Precautions

- Safety Data Sheets (SDS) for individual reagents and kit components are available from ANCHOR Diagnostics (please contact support@anchor-diagnostics.com).
- Use of this product is limited to personnel specially instructed and trained in molecular laboratory techniques.
- Specimens should always be treated as potentially infectious and/or biohazardous material in accordance with safe laboratory procedures.
- Wear protective single-use gloves, a laboratory coat and eye protection when handling specimens or kit components.
- Avoid microbial and nuclease (DNase/RNase) contamination of the specimen and the components of the kit.
- Always use DNase/RNase-free single-use pipette tips with aerosol barriers.
- Use dedicated supplies and equipment, keeping them separate from areas where purified nucleic acids are used for PCR or other downstream applications.
- The user must ensure that nucleic acid extraction and subsequent analysis using the purified nucleic acids are properly controlled (e.g., with appropriate extraction and/or amplification controls).
- Discard sample and assay waste according to your local safety regulations.
- Store reagents protected from direct sunlight.

Instructions for Use – SpeedPURE® Kits

10 ▸ Technical Assistance & Contact Information

For any questions, or if you identify difficulties using our products do not hesitate to contact us:

phone: +49 40 52 471 62 0
email: support@anchor-diagnostics.com

11 ▸ Literature

[1] Mark A. Lever, Andrea Torti, Philip Eickenbusch, Alexander B. Michaud, Tina ŠantlTemkiv, and Bo Barker Jørgensen: A modular method for the extraction of DNA and RNA, and the separation of DNA pools from diverse environmental sample types; Front Microbiol. 2015; 6: 476.

[2] Sonja Berensmeier: Magnetic particles for the separation and purification of nucleic acids; Appl Microbiol Biotechnol 2006; 73:495–504.

[3] Peter E. Vandeventer, Jessica S. Lin, Theodore J. Zwang, Ali Nadim, Malkiat S. Johal, and Angelika Niemz: Multiphasic DNA Adsorption to Silica Surfaces under Varying Buffer, pH, and Ionic Strength Conditions; J Phys Chem B. 2012 May 17; 116(19): 5661–5670.

12 ▶ Troubleshooting Guide

Observation: Low yield or purity of nucleic acids

Possible Cause	Suggestion
Storage of reagents outside defined temperature range (<15°C or >25°C), which may lead to precipitation, inactivation of proteinase K or other effects.	Discard reagents. Store SpeedPURE® kits and buffers under the correct storage conditions (see chapter 6). Repeat the purification procedure using new samples and reagents.
Improper pretreatment of samples.	Ensure samples are prepared according to the instructions in chapter 5.
Frozen samples were not thawed or mixed properly.	Ensure samples are completely thawed and properly mixed before use.
Sample not stored or handled properly before purification of nucleic acids.	Follow instructions for use of sample collection device and/or transportation device.
Eluate not stored properly.	Use eluate for downstream applications directly after sample purification. Store eluates at 2°C to 8°C overnight or at -30°C to -15°C
Inhibitory substances in sample or eluate.	SpeedPURE® Kits are developed to remove inhibitory substances efficiently. For further usage of purified nucleic acids in downstream applications, it is recommended to control amplification efficiency with an appropriate control.

Instructions for Use – SpeedPURE® Kits

Observation: Unprocessed sample

Possible Cause	Suggestion
High sample viscosity or presence of solids in the sample	Ensure samples are prepared according to the instructions in chapter 5. For stool samples, we recommend using the SpeedPURE® Dilution Buffer.
High sample viscosity	For whole blood samples we recommend using the SpeedPURE® Pre-Lysis Buffer.

Observation: Precipitate formation in dilution buffer

Possible Cause	Suggestion
Storage of the buffer at temperatures below 15 °C may lead to precipitation of buffer components.	Incubate SpeedPURE® Dilution Buffer in e.g., water bath at 30°C for 30 minutes. Mix occasionally to resolve precipitates.

13 ▸ Symbols

COMP Components in Kit

CONT Volume in container

LOT Batch code

NUM Number of tubes

QG Quick Guide - Catalog number and version

REF Product - Catalog number

RUO For research use only

UDI Unique Device Identifier

Catalog number and version
Consult Instructions for Use

Important Note

Use by

Contains sufficient reagents for <N> tests

Temperature limits for storage

Manufacturer

GTIN Global Trade Item Number

UFI Unique Formula Identifier (EU) 1272/2008

14 ▸ Trademarks and Disclaimers

SpeedPURE® (ANCHOR Diagnostics GmbH)
Registered names, trademarks, etc. used in this document, even if not specifically marked as such, should not be regarded as unprotected by law.

DIAGNOSTICS

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