ANCHOR

SpeedPURE® Flex Kit (384) (RUO)

1. Turn on the ANCHOR BEAD 32 and log in
2. Prepare Deep Well Plate according to table A
3. **Important! Mix magnetic beads well before pipetting!**
4. Starting from the upper left, only prepare as many rows with reagents as needed for your samples (example shown for 10 samples, Fig. 1)
5. Add 200 µL (or 400 µL, depending on protocol) of sample directly into the Lysis Buffer (Fig. 1; first well, blue). **Do not mix!**
6. Addition of Proteinase K and Internal Control (IC):
   a. IC included as extraction control: add 20 µL of Proteinase K and 9 µL of IC to the sample/Lysis Buffer mix in the first well. **Do not mix!**
   b. IC **not** included as extraction control: add 20 µL of Proteinase K to the sample/ Lysis Buffer mix in the first well. **Do not mix!**
7. Open the ANCHOR BEAD 32 door and place Deep Well Plates into the instrument (Fig. 2) Ensure the Deep Well Plates are securely positioned with the blue lysis buffer on the left
8. Insert the SpeedPURE® Magnetic Rod Covers – always use two rod covers per plate (two or four in total) to ensure proper protection of the magnetic rods (Fig. 3)

Columns	Reagent	Volume [µL]
1, 7	Lysis Buffer (L)	600
2, 8	Wash Buffer I (WI)	600
3, 9	Wash Buffer II (WII)	600
4, 10	Magnetic Beads (B)	80
5, 11	None (empty)	n/a
6, 12	Elution Buffer (E)	90

Table A

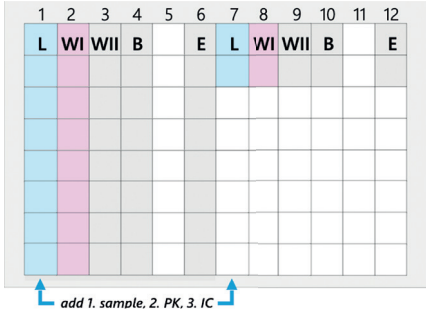

Fig. 1

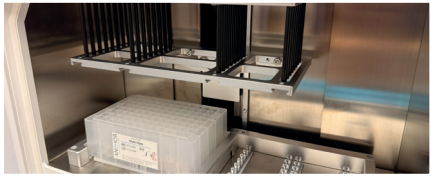

Fig. 2

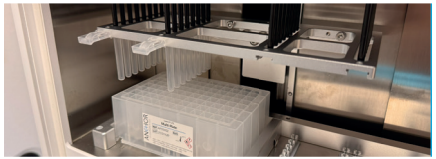

Fig. 3

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9. Ensure all magnetic rod covers click into place and are inserted firmly
10. On the display select Run (upper left corner)
11. Choose the desired protocol (Fig. 4)
    - SpeedPURE_200 for 200 µL sample input
    - SpeedPURE_400 for 400 µL sample input
12. Select Run (lower right corner) to start the sample preparation
13. After the run is complete:
    - Remove and discard magnetic rod covers
    - Remove Deep Well Plates from instrument and collect eluates from columns 6 and 12 (Fig. 5)
    - Discard used Deep Well Plates

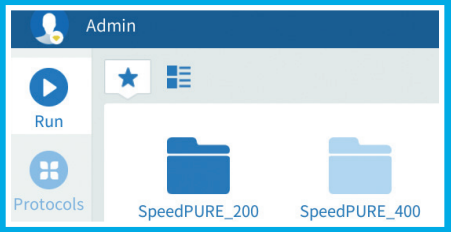

Fig. 4

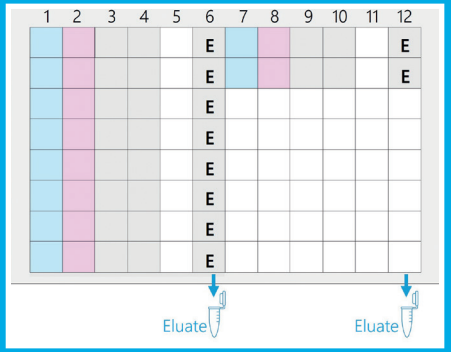

Fig. 5

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