ANCHOR

SpeedPURE® Multi Kit (96) (RUO)

1. Turn on the ANCHOR BEAD 32 and log in
2. Invert each Multi Plate 3 times to resuspend the beads
3. Briefly flick the filled plate downward by hand (3 times) to collect any liquid at the bottom of the wells
4. Remove the seal of each Multi Plate
5. Add 200 µL (or 400 µL, depending on protocol) of sample directly into the Lysis Buffer (Fig. 1; first well, blue). **Do not mix!**
6. Addition of Proteinase K and Internal Control (IC)
   a. IC included as extraction control: add 20 µL of Proteinase K and 9 µL of IC to the sample/Lysis Buffer mix in the first well. **Do not mix!**
   b. IC **not** included as extraction control: add 20 µL of Proteinase K to the sample/Lysis Buffer mix in the first well. **Do not mix!**
7. Open the ANCHOR BEAD 32 door and place Multi Plates into the instrument (Fig. 2) Ensure the Multi Plates are securely positioned with the plate label facing the front of the instrument
8. Insert the SpeedPURE® Magnetic Rod Covers – always use two rod covers per plate (two or four in total) to ensure proper protection of the magnetic rods (Fig. 3)

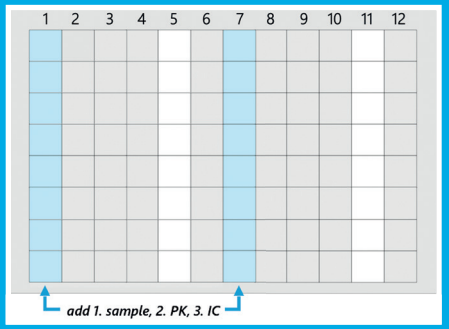

Fig. 1

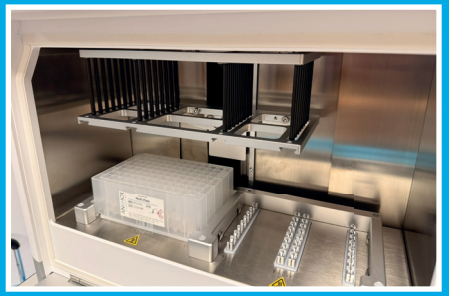
Fig. 2

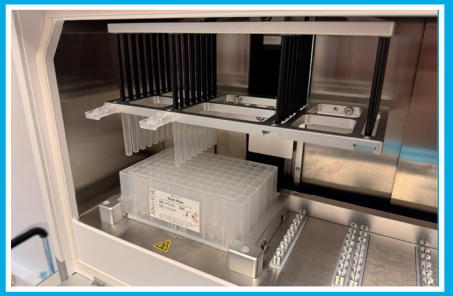
Fig. 3

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9. Ensure all magnetic rod covers click into place and are inserted firmly
10. On the display select Run (upper left corner)
11. Choose the desired protocol (Fig. 4)
    - SpeedPURE_200 for 200 µL sample input
    - SpeedPURE_400 for 400 µL sample input
12. Select Run (lower right corner) to start the sample preparation
13. After the run is complete:
    - Remove and discard magnetic rod covers
    - Remove Multi Plates from instrument and collect eluates from columns 6 and 12 (Fig. 5)
    - Discard used Multi Plates

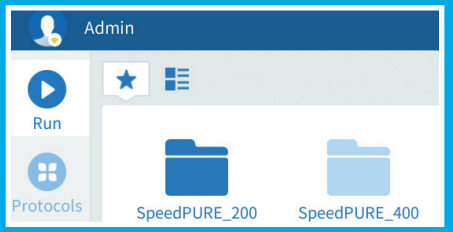

Fig. 4

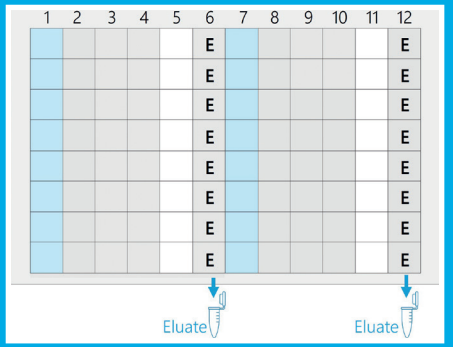

Fig. 5

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